

Magnetic beads retention device for sandwich immunoassay: comparison of off-chip and on-chip antibody incubation

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Abstract We use magnetic microbeads, which are magnetically self-assembled in chains in a microfluidic chip, as reaction substrates to implement two different sandwich immunoassay protocols for the detection of mouse monoclonal target antibodies. The magnetic chains form when the chip is placed in a magnetic field, and are geometrically trapped and accurately positioned in a microchannel with periodically enlarged cross-sections. In the first immunoassay protocol, capture and target antibodies are incubated off-chip, while exposure to the detection antibody is performed on-chip. In the second protocol, the complete immunoassay is fully executed on-chip. In the ‘off-chip incubation–on-chip detection’ protocol, antibodies can be detected down to a concentration of 50 ng/mL in a total assay time of 120 min, while consuming 1.5 mL of target antibody solution. Using the full on-chip protocol, our system is able to detect target antibodies in the range of a few ng/mL in 30 min, using only a few tens of nanoliters of target antibody solution and reagents. The ‘off-chip incubation–on-chip detection’ protocol is also applied for dosing antibodies obtained from the supernatant of a cell culture medium.

Keywords Microfluidics · Immunoassay · ELISA · Antibody · Magnetism · Magnetic microbeads · Self-assembly · Microchip · Nanoparticles

1 Introduction

The miniaturization of analytical chemistry systems has become a major challenge in the research community since the introduction of the micro total analysis system (μ -TAS) in 1990 (Manz et al. 1990). Immunoassay analyses (for example the Enzyme-Linked Immunosorbent Assay (ELISA), an analytical technique widely used in clinical diagnostics, cancer research, and food safety or environmental analysis) use the sensitivity and specificity of highly stable interactions between antigens (Ag) and their specific antibodies (Ab). An immunoassay analysis is conventionally performed in microtiter plates, and consumes typically about hundreds of μ L of sample and reagent solutions, for an assay time of hours due to mass-transport limitations. In microfluidics, the small reaction volume and microchannel dimensions permit a strong analysis time reduction, while consuming minute quantities of samples and reagents (typically a few μ L). However, microfluidic integrated systems developed for immunoassays demand additional steps for a perfect control of microchannel surface chemistry, to allow an efficient and reproducible Ab immobilization. This drawback can be circumvented by using functionalized microbeads in a fluidic chip. Microbeads are available at different sizes, present the advantage of a large surface-to-volume ratio, and offer a strong flexibility due to a wide range of available surface chemistries. Their use as substrate for microfluidic immunoassays was investigated by elaborating microbead retention strategies at desired positions within a microchannel, using for example a microdam (Sato et al. 2001) or micropillar cage (Andersson et al. 2000). When microbeads incorporate a magnetic core, they can be easily manipulated using external magnetic fields, explaining why they are increasingly used as a solid phase for Ab capture and as carriers of

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captured Ag (Gijs 2004). Their retention at the microchannel wall was realized using macroscopic permanent magnets (Hayes et al. 2001), current-activated coils (Choi et al. 2002) and wires (Siegel et al. 2006), or magnetic structures (Deng et al. 2002; Lund-Olesen et al. 2007) incorporated on the microchip. Mobile permanent magnets were also used for magnetic microbeads' manipulation in a microfluidic chip incorporating a dual channel network, 460 nL reaction chambers and valve-activated fluidic bridges (Herrmann et al. 2006, 2007). Recently, the manipulation and retention of magnetic microbeads over the complete cross-section of a microchannel was proposed using a focused magnetic field (Rida and Gijs 2004; Slovakova et al. 2005; Smistrup et al. 2005; Bronzeau and Pamme 2008; Peyman et al. 2008), or by trapping self-assembled magnetic chains geometrically in a periodically structured microchannel (Lacharme et al. 2008a, b). In these systems, the intense liquid-magnetic microbead interactions lead to a highly efficient active mixing or target analyte capture.

In this paper, we present and compare two protocols to perform on-chip immunoassays using our magnetic microbead retention device. The magnetic microbeads are incubated with capture Ab (c-Ab) and target monoclonal Ab (t-mAb) either off-chip or directly on-chip (t-mAb is used as target antigen in our sandwich immunoassay), while exposure to the detection Ab (d-Ab) is always performed on-chip. The off-chip incubation–on-chip detection protocol extends the use of our chip to magnetic microbead-based analytical methods involving classical bench-top Ab analysis procedures. A controlled amount of magnetic microbeads is self-assembled in magnetic chains that are accurately retained in the large sections of a periodically enlarged microchannel, by simply placing the chip in an external homogeneous magnetic field. The self-assembled magnetic chains strongly interact with analytes and d-Ab from the perfusing microfluidic flows. We compare the performance of both immunoassay protocols and, as additional application example, demonstrate the dosing of t-mAb produced by a hybridoma cell culture.

2 Microsystem design and fabrication

The schematic layout of the microchip showing the periodical enlargement of the microchannel is presented in Fig. 1a. It consists of a 20- μm -wide microchannel with periodical sections of 40 μm (of 5 μm length) separated by 10 μm . 130 sections of this type are realized over a total distance of 2 mm. The microchips are microfabricated using a dry-etching technology (Pavus 2004), to obtain a vertical microchannel wall profile. Briefly, a 2- μm -thick layer of amorphous silicon (a-Si) is deposited on a 4-inch Pyrex wafer, following by spinning, exposure, and development of a positive photoresist

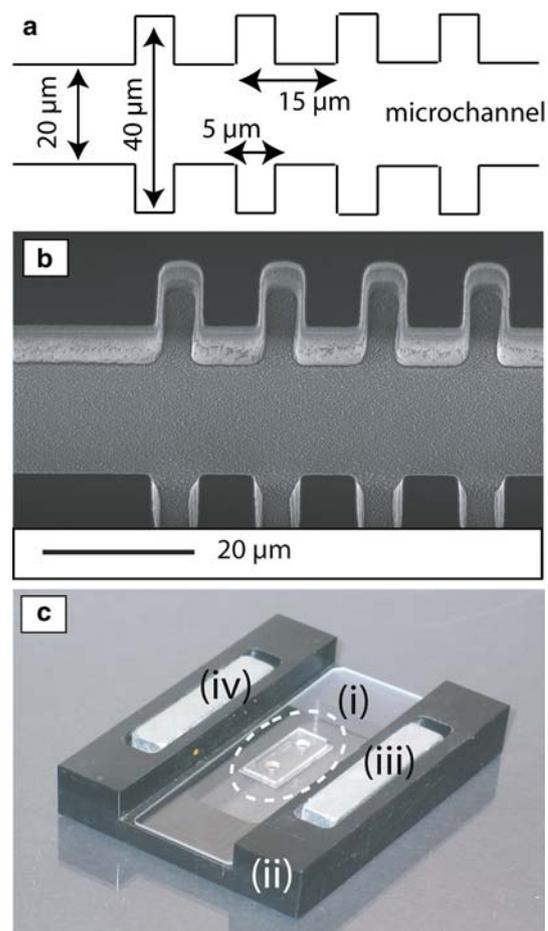


Fig. 1 **a** Schematic layout of the 20- μm -wide microchannel showing the periodically enlarged sections of 40 μm . **b** SEM picture of the microchannel showing the periodically varying channel width. **c** Picture of the microfluidic chip in its socket. The chip consists of a 8- μm -deep plasma-etched channel in a Pyrex substrate that is sealed with a 2.5 mm thick Polydimethylsiloxane (PDMS) cover (i). The latter is pinched with two access holes and placed on the chip socket (ii) that contains two permanent magnets (iii, iv) generating a homogeneous magnetic field

using a standard photolithography process. The a-Si is dry etched using a deep reactive ion-etching (DRIE) technique, and the remaining photoresist is stripped. Then, the Pyrex wafer is DRIE etched to a depth of 8 μm , followed by removal of the a-Si protective mask in a 40% KOH bath, heated at 60°C during ~ 1 h. Finally, the wafer is cut in individual microchips. Figure 1b shows a scanning electron microscopy (SEM) image of the microchannel. Finally, the microchip is loosely covered with a 2 mm thick polydimethylsiloxane (PDMS) cover, which is pinched with two access holes forming two reservoirs (reservoir A and B). Figure 1c shows a picture of the microchip (i) on its holder used during the experiments, where it is placed at the center of a poly-oxymethylene support (ii), and flanked by two permanent magnets (iii and iv) for the generation of the permanent homogeneous magnetic field (Rida et al. 2003).

3 Magnetic bead retention principle

A solution of 500 nm diameter magnetic microbeads is introduced into the microchannel. When a homogeneous magnetic field is applied perpendicularly to the microchannel axis, the magnetic microbeads self-assemble in magnetic chains (Doyle et al. 2002) in the microchannel’s large cross-sections, thereby decreasing their total magnetic dipolar energy. Figure 2a is a schematic drawing of the system where a flow is applied through a magnetic chain, exerting a drag force F_{drag} which displaces the central part of the chain over a distance X . The saturation magnetization of the microbeads is $M_{\text{sat}} = 8 \times 10^4$ A/m. Then, the maximal magnetic moment of one bead $m_{\text{bead}} = (4/3)\pi R^3 M_{\text{sat}} = 5.23 \times 10^{-15}$ Am², where R is the bead radius (250 nm). We measure with a miniaturized Hall probe a magnetic induction of 120 mT originating from the two permanent magnets at the center of the chip holder, where the microchannel is inserted during experiments. This magnetic induction is large enough to saturate the magnetic moment of the microbeads. Assuming that all the magnetic microbeads have reached their magnetic saturation (due to the permanent magnetic field) and are individual dipoles oriented in the external magnetic field

direction, the magnetic dipole energy of a pearl-like chain of n microbeads is given by (Helseth 2005)

$$E_{\text{chain}} = \sum_{i=0}^{i=n} \sum_{j>i}^n \frac{\mu_0}{4\pi} \frac{1}{r_{ij}^3} m_{\text{bead}}^2 \times (1 - \cos^2 \Theta_{ij}) \tag{1}$$

where μ_0 is the magnetic permeability of vacuum, r_{ij} the distance between the microbeads with index i and j , and Θ_{ij} the angle between the external magnetic field direction and the center-to-center vector of microbeads with index i and j , respectively. When exposed to the flow, the viscous drag force acting on the central part of 80- microbead pearl-like magnetic chain is approximated by the Stokes drag force:

$$F_{\text{drag}} = 6C\pi\eta Rv \tag{2}$$

where C is the number of magnetic microbeads in the center part of the chain (40 for a 80-magnetic microbead pearl-like chain trapped in a 40 μm enlarged microchannel section), η the liquid viscosity and v the velocity of the liquid with respect to the magnetic microbeads. Figure 2b presents the magnetic dipolar energy E_{chain} for a chain of 80 microbeads calculated using Eq. 1, as a function of X . The magnetic energy of the chain increases with X , as the individual microbead center-to-center distances increase. This energy variation leads to a magnetic dipolar force given by

$$F_{\text{chain}} = -\frac{\partial E_{\text{chain}}}{\partial X} \tag{3}$$

Figure 2c presents the variation of the magnetic dipolar force F_{chain} as a function of X calculated using Eq. 3. A maximum force of ~ 270 pN is calculated for a distance $X = 250$ nm. When the drag force is superior to the magnetic force ($F_{\text{drag}} > F_{\text{chain}}$), the central part of the magnetic chain is no longer retained and the chain breaks in three parts. Equalizing F_{drag} and F_{chain} lead to a critical mean fluid velocity for chain rupture of $v_c = 1.4$ mm/s. During our experiment, we typically use a maximum flow velocity at the microchannel center of 160 $\mu\text{m/s}$ (i.e., a flow rate of 0.034 nL/s). Then, no chain breaking is observed during experiments, and the self-assembled magnetic chains stay retained and are accurately positioned at the entire cross-section of microchannel.

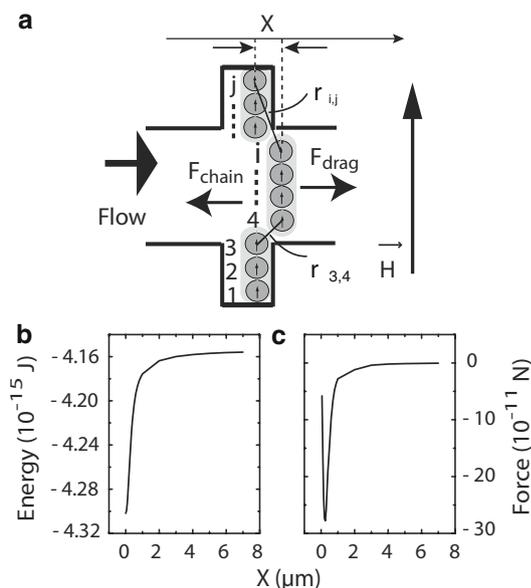


Fig. 2 a Schematic layout of the microchannel with periodically varying channel width. In a magnetic field H , the magnetic microbeads self-assemble into chains that stay trapped in the large cross-sections. When a fluid flow is applied, the viscous drag force displaces the center part of the microbead chains. $r_{i,j}$ is the distance between the nanoparticles with index i and j , respectively. b Magnetic dipolar energy E_{chain} of a chain consisting of 80 magnetic microbeads as a function of the displacement X induced by the flow. c Magnetic retention force F_{chain} of a chain consisting of 80 microbeads as a function of X

4 Experimental

All c-Ab and d-Ab solutions are prepared by diluting the desired amount of Ab in phosphate buffer saline–bovine serum albumine (PBS–BSA) solution. (PBS–BSA) solution is prepared by diluting 1% (w/w) BSA (obtained from Sigma-Aldrich, product no. A7030) in PBS (obtained from Sigma-Aldrich, product no. P5493) solution (pH 7.4). In this study, all Ab used are immunoglobulin G (IgG). The

c-Ab molecules used for both on- and off-chip immunoassay are rabbit biotinylated anti-mouse IgG (obtained from Dako, product no. E0354). The direct detection Ab (dd-Ab) molecules used for performing a controlling direct immunoassay are Cy3-conjugated polyclonal goat anti-rabbit IgG (obtained from Bioconcept, product no. 601-104-122). The sandwich detection Ab (sd-Ab) molecules used for the sandwich immunoassay are Cy3-conjugated polyclonal goat anti-mouse IgG (obtained from BioConcept, product no. 610-104-121). The t-mAb molecules used for the sandwich immunoassay are normal mouse IgG (obtained from Invitrogen, product no. 10400C). The t-mAb solutions are prepared by diluting 39 $\mu\text{g}/\text{mL}$ initial concentrated solution in cell culture medium (obtained from Invitrogen) supplemented with 5% fetal calf serum (FCS, obtained from Invitrogen). Streptavidin coated magnetic microbeads (MasterBeads Streptavidin obtained from Ademtech, product no. 03153) are used as magnetic microbeads for both the on- and off-chip immunoassay. Immobilization Buffer (obtained from Ademtech, product number no. 10301) is used as washing buffer (WB), according to the supplier's recommendations. For both protocols, before introducing magnetic microbeads in the microchannel, we first pour a surface-blocking solution through the microchip to avoid aspecific Ab adsorption on the glass channel wall and on the PDMS cover. A volume of 15 μL of PBS-BSA solution is introduced in one reservoir (reservoir A), after which the solution fills the entire microchannel by capillary forces within a few seconds. Then, the second reservoir (reservoir B) is also filled with 15 μL of PBS-BSA solution. After 5 min, both reservoirs are emptied by withdrawing solutions using a micropipette, and rinsed with 15 μL of WB.

The 'off-chip incubation–on-chip detection' protocol is illustrated in Fig. 3. In a first step, 10 μL of magnetic microbead solution is resuspended in 10 μL of WB, to avoid magnetic beads' clustering or reaction vial surface adsorption before c-Ab incubation. The magnetic microbeads are then incubated for 30 min under gentle agitation with 166 μL of c-Ab diluted at 455 ng/mL (Fig. 3i). Using this c-Ab solution concentration, the size of magnetic microbead agglomerates formed because of the link between the surface streptavidin molecules and the c-Ab is limited, allowing the functionalized magnetic microbeads to be easily introduced in microchannel. Then, the magnetic microbeads are washed with 500 μL of WB and resuspended in 10 μL of WB. In a second step, the microbeads are incubated with 500 μL of t-mAb solution (Fig. 3ii) at different concentrations (from 50 to 200 ng/mL) for 60 min under gentle agitation. Then, the microbeads are washed with 500 μL of WB and resuspended in 10 μL of WB, avoiding further magnetic bead adsorption to the microchannel surface. The final magnetic microbead concentration is of 10 mg/mL (Fig. 3iii). A volume of 2 μL of the magnetic microbead solution is introduced in reservoir A and flushed into the microchannel by applying a short over-pressure in the reservoir. Then, the microchip is placed on its holder supporting the permanent magnets. All magnetic chains are self-assembled after ~ 10 s (Fig. 3iv). The magnetic microbead solution remaining in reservoir A is withdrawn from the microchip with a micropipette, and reservoir A is filled with 4 μL of PBS-BSA solution. To execute the on-chip detection, we fill reservoir B with 15 μL of sd-Ab solution at 5 $\mu\text{g}/\text{mL}$ (Fig. 3v). The liquid volume difference between reservoirs A and B provides the pressure to generate a liquid flow velocity of 160 $\mu\text{m}/\text{s}$ at

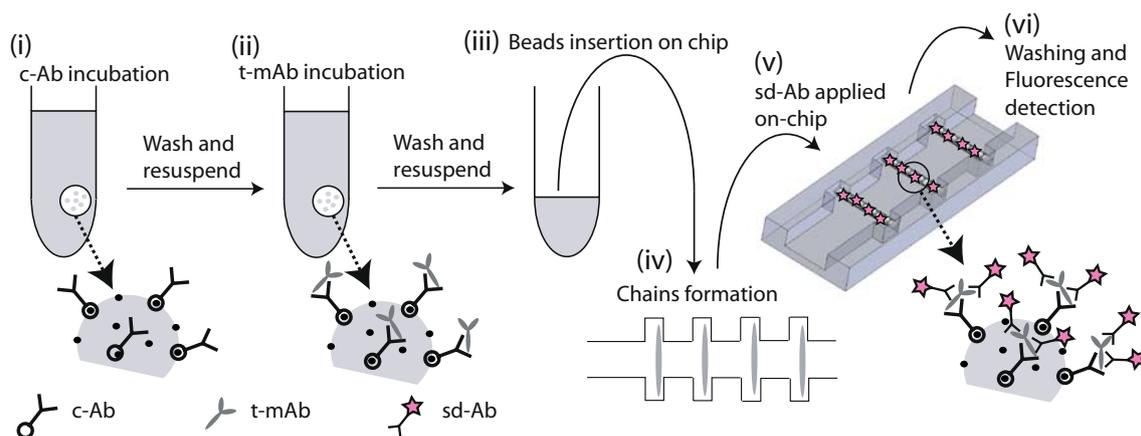


Fig. 3 Illustration of the 'off-chip incubation–on-chip detection' protocol. *i* The magnetic microbead solution is incubated with capture antibodies (*c-Ab*). *ii* The microbeads are incubated with target monoclonal antibodies (*t-mAb*) solution at different concentrations under gentle agitation. *iii* The magnetic microbeads are

introduced in the microchip, where (*iv*) the magnetic chains self-assemble. *v* Flow of the sandwich detection antibodies (*sd-Ab*) solution through the magnetic chains. *vi* After a washing step, the fluorescence of the chains can be detected

Table 1 Summary of the experimental parameters used for the three experimental protocols

Experimental step	Total number of beads	Ab solution volume (nL)	Ab solution concentration (ng/mL)	Total number of Ab molecules	Average number of Ab molecules per bead
Off-chip incubation–on-chip detection					
Incubation of c-Ab (off-chip)	7.63×10^8	166×10^3	455	3.02×10^{11}	396
Incubation of t-Ag (off-chip)	7.63×10^8	500×10^3	From 50 to 200	From 1×10^{11} to 6×10^{11}	From 131 to 786
Application of sd-Ab (on-chip)	29,000	15.3	5,000	3.24×10^8	11,066
Full on-chip direct immunoassay					
Application of c-Ab	29,000	6.12	5,000	1.22×10^8	4,167
Application of dd-Ab	29,000	10.2	5,000	2.04×10^8	6,967
Full on-chip sandwich immunoassay					
Application of c-Ab	29,000	6.12	5,000	1.22×10^8	4,167
Application of t-mAb	29,000	30.6	From 1 to 200	From 1.22×10^4 to 2.45×10^6	From 4 to 836
Application of sd-Ab	29,000	10.2	5,000	3.24×10^8	11,066

the microchannel center (Shui et al. 2007), without the need of external complex tubing or pumping system. The sd-Ab solution is flown through the microchannel for 7.5 min, to ensure an excess amount of d-Ab with respect to the t-mAb. Then, reservoir B is rapidly rinsed (with WB), before application during 3 min of WB through the microchannel by filling reservoir B with 15 μ L of WB solution. Finally, reservoir B is filled with 15 μ L of PBS–BSA solution during fluorescence detection (Fig. 3vi).

The full on-chip immunoassay protocol has been previously described in detail (Lacharme et al. 2008a, b). Briefly, 2 μ L of 10 mg/mL magnetic microbead solution (resuspended in WB to avoid magnetic beads adsorption to the microchannel wall surface) is introduced in reservoir A (before c-Ab coating), and flushed into the microchannel by applying a short over-pressure in reservoir A. The microchip is placed on its holder containing the permanent magnets and chains form. Excess of magnetic microbead solution is withdrawn from reservoir A using a micropipette and replaced by 4 μ L of PBS–BSA solution. All other fluid operations are performed by filling and withdrawing 15 μ L of different solutions from reservoir B. The on-chip direct control immunoassay is performed by subsequently letting flow through the microchannel (from reservoir B to reservoir A) solutions of (1) c-Ab at 5 μ g/mL for 3 min and (2) dd-Ab at 5 μ g/mL for 5 min. The sandwich immunoassay is performed by subsequently letting flow through the microchannel (from reservoir B to reservoir A) solutions of (1) c-Ab at 5 μ g/mL (in PBS only solution) for 3 min, (2) t-mAb solutions at different concentrations (from 1 to 200 ng/mL) for 15 min and (3) sd-Ab at 5 μ g/mL for 5 min. All steps (during both the direct and sandwich immunoassay) are separated by a fast-rinsing (\sim 10 s) and 3 min washing step using 15 μ L of WB solution introduced in reservoir B. Finally, the reservoir B is filled with 15 μ L

of PBS–BSA solution during fluorescence detection. Table 1 summarizes the experimental parameters (total number of magnetic microbeads used, Ab solution volume and Ab concentration, total number of Ab molecules and average number of Ab molecules per bead) used for the three experimental protocols.

For visualization, image acquisition, and fluorescence detection, we use a CCD camera ORCA-C4742-95ER (Hamamatsu) mounted on an inverted microscope Axiovert S100 (Zeiss), with a 100 \times objective (LD EC Epiplan-Neofluar, numerical aperture 0.75), a 40 \times objective (LD Achromplan, numerical aperture 0.5) and a 10 \times objective (Epiplan, numerical aperture 0.2). The microscope is equipped with a Mercury short arc lamp HBO (Osram) and the appropriate filter set for Cy3 dye (filter set no. 20, Zeiss). The excitation, dichroic mirror, and emission wavelengths for this filter set are 446–512, 560 and 575–640 nm, respectively. The exposure time is 2 s and AquaCosmos software (Hamamatsu) is used for the fluorescence image analysis and for correcting the data for the background detection signal. Typically, an area of $170 \times 20 \mu\text{m}^2$ with nine consecutive magnetic chains is analyzed for the immunoassay experiments.

5 Results and discussion

Figure 4a shows an optical image (taken with the 100 \times objective) of five self-assembled magnetic chains before c-Ab coating. During experiments, the magnetic microbeads first assemble in primary linear chains in the narrow and wide sections of the microchannel before constituting 40- μ m-long magnetic chains that stay trapped in the 40- μ m-large section of the microchannel. Thereby, the chains are more complex than a pearl-like chain. We

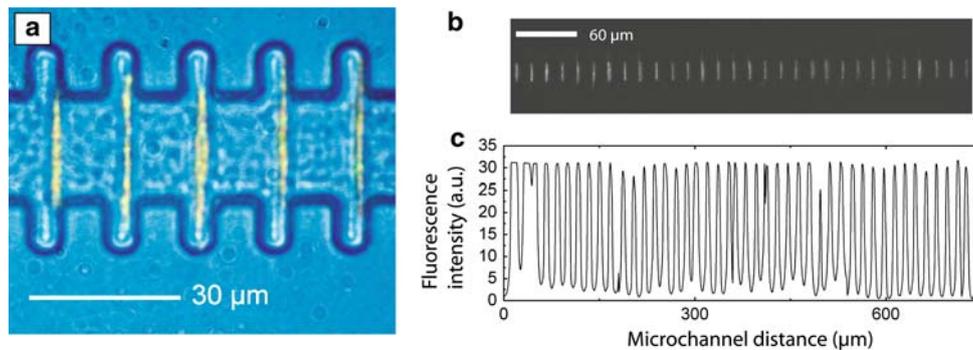


Fig. 4 **a** Optical image of five self-assembled magnetic chains. **b** Fluorescence microscopic image of 30 consecutive self-assembled magnetic microbead chains after the realization of the direct control

immunoassay. **c** Fluorescence intensity along the microchannel axis due to the formation of the two-Ab fluorescent immunocomplexes on the magnetic microbeads, as derived from images like shown in **b**

observe in Fig. 4a linear chains of $\sim 1.5 \mu\text{m}$ diameter with local small imperfections. To estimate the number of magnetic microbeads constituting a chain, we calculate the number of magnetic microbeads initially present in one trapping unit of the microchannel (a volume of 3.2×10^{-9} mL consisting of one large and one narrow section of the microchannel). With an initial magnetic microbead concentration of 10 mg/mL, we conclude that a single magnetic chain is constituted of approximately 240 magnetic microbeads. Figure 4b shows a fluorescence microscopy image of ~ 30 chains in the microchannel obtained after completion of the full on-chip direct control immunoassay. The two-Ab fluorescent immunocomplexes (supramolecules constituted by c-Ab and dd-Ab) are clearly located at the surface of the chains, indicating the high capture efficiency of the magnetic microbeads in our system. Figure 4c shows the fluorescence intensity along the microchannel axis due to the binding of the two-Ab fluorescent immunocomplexes on magnetic microbeads, as derived from images like that shown in Fig. 4b. Each peak corresponds to a single fluorescent magnetic chain. In this direct control immunoassay, all chains that are periodically trapped over the 2-mm-long microchannel (representing a total number of 120 chains) show strong fluorescence intensity. The large number of c-Ab linked to the surface of the magnetic microbeads are equivalently distributed over the 240 microbeads per chain \times 120 (chains) \cong 29,000 magnetic microbeads in the microchannel. During flowing of the c-Ab solution, 1.22×10^8 c-Ab molecules are introduced in the microchannel. On average, 4,167 c-Ab are linked at the surface of each microbead in the microchannel, and this by only consuming 6.12 nL of c-Ab solution.

Figure 5a shows an optical microscopy image of the self-assembled magnetic chains in the microchannel after off-chip c-Ab and t-mAb incubation. The chains are accurately positioned in the geometrical traps. However, their size and homogeneity present slight variations, due to the off-chip incubation protocol. Indeed, when off-chip

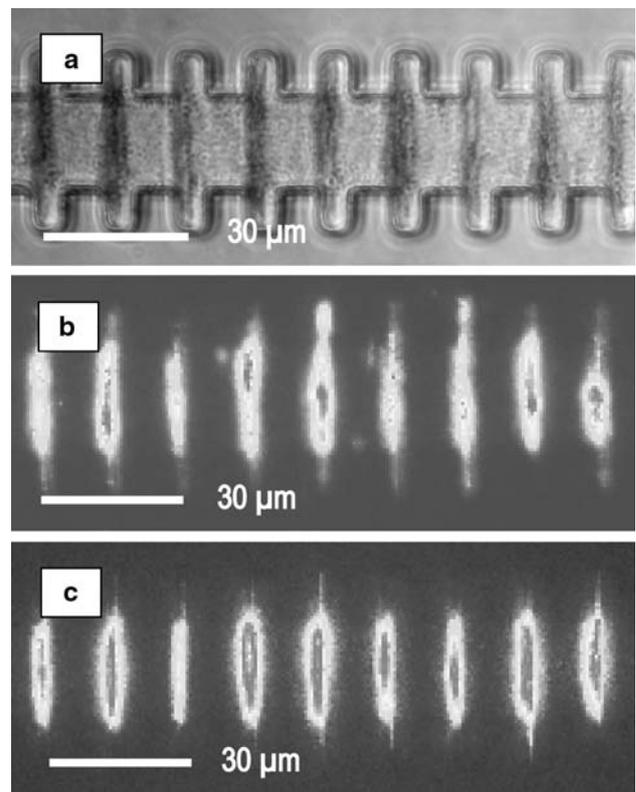


Fig. 5 **a** Optical image of the self-assembled magnetic chains after the off-chip c-Ab and t-mAb incubation. **b** Fluorescence image of the self-assembled magnetic chains in the microchannel after completion of the 'off-chip incubation-on-chip detection' sandwich immunoassay. **c** Fluorescence image obtained after completion of the full on-chip sandwich immunoassay

incubated, the magnetic microbeads are linked by c-Ab and form small microbead clusters. Once introduced in the microchannel, the clusters are embedded in the self-assembled magnetic chains during their formation and create chain irregularities. Figure 5b shows a fluorescence image of the self-assembled magnetic chains in the microchannel after completion of the 'off-chip incubation-on-chip detection'

sandwich immunoassay. The three-Ab fluorescent immunocomplexes (supramolecules constituted by c-Ab, t-mAb and sd-Ab) are clearly located on the magnetic chains. The fluorescence signal presents variations along the chain axis due to the chain irregularities. This phenomenon is not observed in the full on-chip sandwich immunoassay (where we introduce the magnetic microbeads in the microchannel before c-Ab coating), because the microbeads behave as independent particles which self-assemble only due to magnetic interactions when exposed to the external magnetic field. For comparison, Fig. 5c shows a fluorescence image obtained after completion of the full on-chip sandwich immunoassay. The chains are accurately positioned in the geometrical traps and are regular in size, while the fluorescent signal is homogeneous along the chain axis.

Figure 6a shows the fluorescence intensity of nine magnetic chains along a section of the microchannel, as derived from an image like shown in Fig. 5b, after completion of the ‘off-chip incubation–on-chip detection’ immunoassay. In a sandwich immunoassay, the limiting factor for the three-Ab immunocomplex formation is the

number of t-mAb. During the off-chip incubation, c-Ab and t-mAb are homogeneously distributed over the magnetic microbead surface. The total fluorescence signal averages out chain-to-chain intensity fluctuation. The integration over all fluorescent chains on the image is proportional to the total number of t-mAb captured on the magnetic microbeads. Figure 6b shows the total integrated fluorescence as a function of the t-mAb concentration, obtained from curves like shown in Fig. 6a. Although the average number of c-Ab per magnetic microbead is lower than the average number of t-mAb per bead for solution concentrations above 150 ng/mL (see Table 1), the signal increases linearly for t-mAb concentrations between 50 and 200 ng/mL. This indicates that a portion of t-mAb molecules are probably not captured, when incubated off-chip. The ‘off-chip incubation–on-chip detection’ protocol is also applied to 500 μ L of the supernatant of a cell culture to measure the t-mAb concentration in a non-purified sample solution. The cell culture is prepared by conventional hybridoma technology (Plessers et al. 1990) in a conditioned cell culture medium to produce t-mAb (5D10

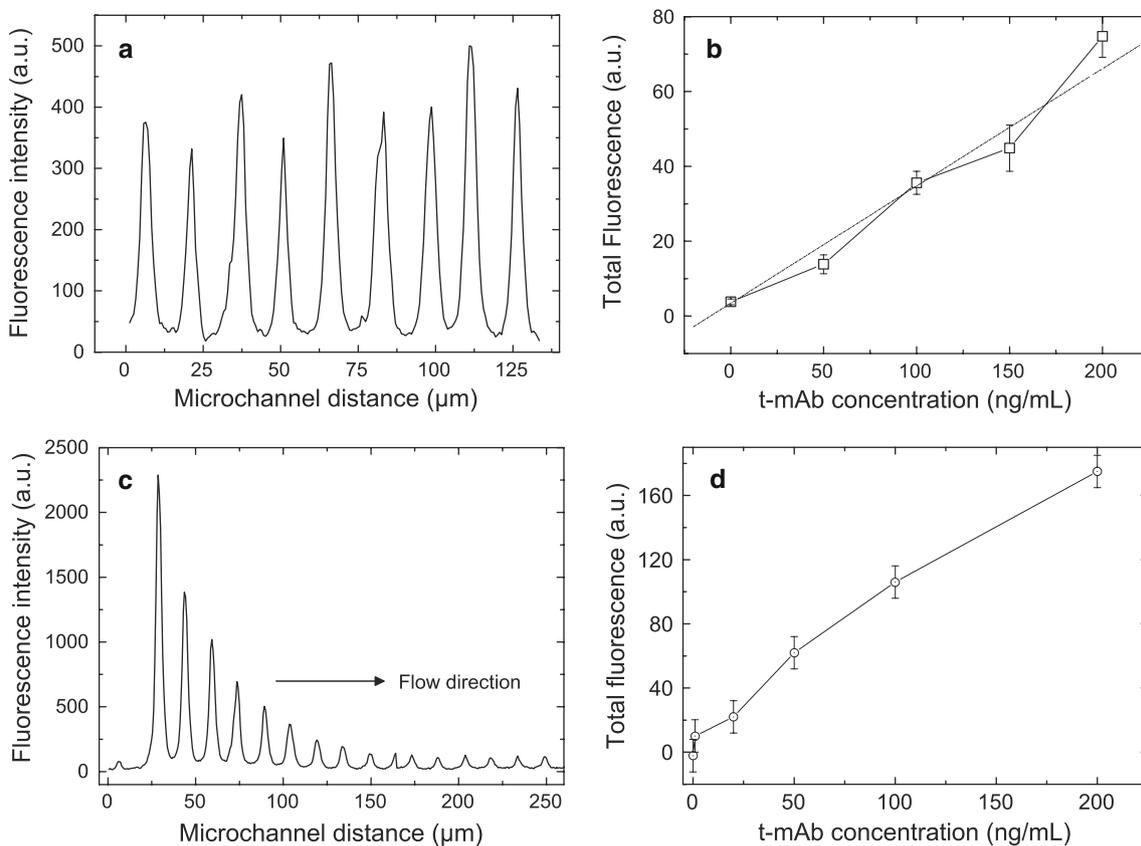


Fig. 6 **a** Fluorescence intensity along a section of the microchannel after completion of the ‘off-chip incubation–on-chip detection’ sandwich immunoassay. **b** Total fluorescence as a function of the t-mAb concentration for the ‘off-chip incubation–on-chip detection’

sandwich immunoassay. **c** Fluorescence intensity along the microchannel after the completion of the full on-chip sandwich immunoassay. **d** Total fluorescence as a function of the t-mAb concentration for the full on-chip sandwich immunoassay

Ab directed against the human breast cancer cell line MCF-7). Using the total fluorescence response curve of Fig. 6b, we find a concentration of 113 ± 7 ng/mL. For comparison, this concentration is measured to be ~ 125 ng/mL by a classical ELISA test (in a total assay time of 3.5 h).

Figure 6c shows the fluorescence intensity of the first 16 chains along the microchannel after the completion of the full on-chip sandwich immunoassay, as derived from images like the one shown in Fig. 5c. The chains that are more upstream positioned in the microchannel show the highest fluorescence intensity, which is an indication of the high capture efficiency. Integrating overall fluorescent chains on the complete microchannel provides the total number of t-mAb captured by all chains. Figure 6d shows the total fluorescence intensity as a function of the t-mAb concentration, obtained from curves like the one shown in Fig. 6c. As expected, the signal increases with the t-mAb concentration.

It is clear from Fig. 6c that, during the full on-chip sandwich immunoassay, the magnetic chains gradually deplete the t-mAb concentration downstream of the flow. Consequently, the t-mAb molecules are not homogeneously distributed over all chains. In the absence of Ab saturation effects on the microbeads, the t-mAb concentration, exposing the N th chain in the microchannel and proportional to its fluorescence intensity, can be written as

$$C_N = C_0 \times (1 - P)^N \quad (4)$$

where C_0 is the initial concentration of t-mAb and P (< 1) the capture probability of the chains. Figure 7 presents the experimental maximal fluorescent intensity extracted from

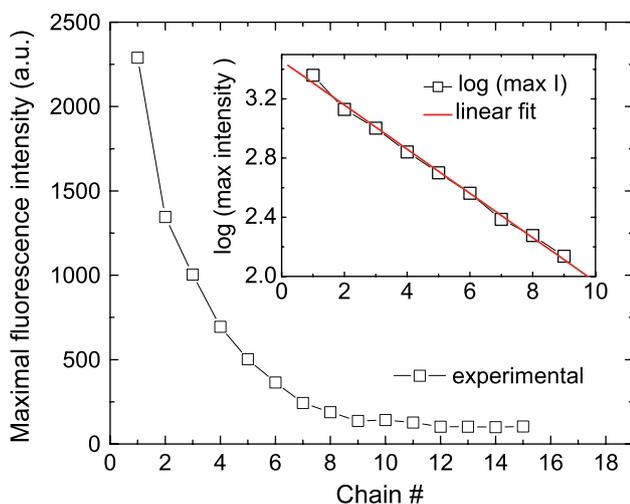


Fig. 7 Experimental maximal fluorescent intensity extracted from Fig. 6c for chains # 1–15. The maximal fluorescent intensity follows an exponential decrease as a function of the chain number. The *insert* shows a single log scale plot of the maximal fluorescent intensity versus chain number for chains # 1–9 (full on-chip sandwich immunoassay)

Fig. 6c for chains # 1–15. As predicted by Eq. 4, the maximal fluorescent intensity follows an exponential decrease as a function of the chain number. The experimental capture probability of a chain can be extracted from experimental data by plotting on a single log scale the maximal fluorescent intensity versus chain number (see insert of Fig. 7).

$$\log\left(\frac{C_N}{C_0}\right) = N \log(1 - P) \quad (5)$$

For our flow condition (flow rate of 0.034 nL/s), the experimental capture probability of a chain determined using Eq. 5 is found to be $P = 0.29$ (i.e., 29% of all t-mAb reaching a microchannel elementary volume, in which one chain is located, will be captured, while the rest continues to advance through the microchannel and will be captured by further chains). Both reaction kinetics and diffusion effects have an influence on the capture probability. Due to diffusion, a minimum number of chains are needed to capture all t-mAb from the flow, even for instantaneous immunoreactions at the microbead surface. Diffusing over a distance corresponding to half of the microchannel height $h/2 = 4$ μm , with a diffusion coefficient for t-mAb of $D = 40 \times 10^{-8}$ cm^2/s , typically takes $h^2/(4D) \approx 0.4$ s. Using an average flow speed of 100 $\mu\text{m}/\text{s}$, we find that, at minimum, the flow needs to pass over a length of 40 μm before all t-mAb molecules have diffused to a magnetic chain. This corresponds to a minimum number of three chains that will show fluorescence, in qualitative agreement with the experimental results. When the flow rate decreases down to a flow velocity of 37 $\mu\text{m}/\text{s}$, the time allowed for a t-mAb molecule to diffuse to the chain surface will increase, and only two chains would be sufficient to capture all t-mAb. Decreasing further the flow rate down to a value below the typical diffusion speed (10 $\mu\text{m}/\text{s}$) will result in the capture of all t-mAb by the first chain of the system.

6 Conclusion

We have demonstrated two protocols to perform on-chip sandwich immunoassays, using magnetic microbeads that are self-assembled in magnetic chains as reaction substrate. When incubated off-chip, t-mAb are distributed over all the magnetic microbead surfaces, and the fluorescence signal obtained reaches a similar level for all chains in the microchannel. This protocol demonstrates the extension of the use of our magnetic microbead retention device to applications involving off-chip immunoreactions with magnetic microbeads. When the t-mAb molecules are captured from a flow in the microchannel, the magnetic chains gradually deplete the t-mAb concentration downstream of the flow, and the t-mAb molecules are unequally

distributed on all chains. From the spatial dependence of the fluorescence intensity, we calculate a magnetic chain capture probability of 29%, primarily due to the finite diffusion times needed for the t-mAb to reach a magnetic chain. In the ‘off-chip incubation–on-chip detection’ protocol, antibodies can be detected down to a concentration of 50 ng/mL in a total assay time of 120 min, while consuming 1.5 mL of target antibody solution. Using the full on-chip protocol, our system is able to detect target antibodies down to a few ng/mL in 30 min, and using only a few tens of nanoliters of target antibody solution and reagents. Our work shows that positioning magnetic microbeads in chains across a microchannel results in a very strong particle-liquid interaction and strongly enhances the fluid perfusion through the magnetic structures. Our results suggest a high potential for immunoassays performed on-chip in our simple and versatile system, consuming lower reagent volumes and strongly reducing experimental time and costs. Moreover, we presume that the principle of magnetic chains’ retention in a microfluidic flow, and incubation of the magnetic beads with analytes, either off- or on-chip, can equally well be used for other bio-analytical or catalytic applications.

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